

Original Article**Antibiotic sensitivity patterns of uropathogens in hospitalized patients at Teaching Hospital Jaffna**
¹Thilukshikka K, ¹Anuruddha AN, ¹Spelman Croos MV, ¹Erandi R, ¹Pravina S, ¹Kumar R, ¹Balagobi B, ²Rajanthi R, ²Ambalavanar V¹Faculty of Medicine, University of Jaffna,²Teaching Hospital Jaffna**Abstract**

Indiscriminate use of antibiotics has led to the emergence of antibiotic resistance in hospital settings. Awareness of local antimicrobial resistance patterns is essential for prudent empirical therapy of urinary tract infections.

This study describes the uropathogens isolated, their antibiotic sensitivity patterns and associated factors in adult inpatients with a positive urine culture at Teaching Hospital Jaffna.

All positive urine culture reports and relevant request forms of adult inpatients (≥ 18 years) investigated at the Microbiological Unit of Teaching Hospital Jaffna during a three-month period (October 1st to December 31st 2020) were analyzed retrospectively with SPSS v27. Standard descriptive statistics and the chi square test were used (critical value 0.05).

Data were extracted from 426 culture reports. Mean age of the sample was 53.2 years (SD 19.9); 47.2% (n=201) of the reports belonged to patients ≥ 60 years and 60.1% (n=256) were of females. Antibiotics prescribed prior to culture were documented in 183 (43%) reports. The most commonly prescribed empirical antibiotic was co-amoxiclav (24%, n=183). Coliforms were the commonest isolate (63.4%, n=270) and showed resistance to several commonly prescribed antibiotics; antibiotic sensitivity was relatively low to ampicillin (9.5%), ceftriaxone (40%) and amoxicillin (48.1%); highest susceptibility was to meropenem (87.6%). Age group and gender were significantly associated with the type of uropathogen isolated ($p \leq 0.05$).

Prior antibiotic therapy was common among inpatients with urinary tract infection at the Teaching Hospital

Jaffna. Sensitivity patterns suggest that antibiotic resistance is a major concern. Empirical therapy needs to be guided by institutional policies and local sensitivity patterns.

Keywords: Uropathogens, Urinary tract infection, Antimicrobial resistance, Antibiotic therapy, Coliforms

Introduction

Urinary tract infection (UTI), which refers to the invasion and growth of microorganisms in the urinary tract, is a common problem in clinical practice (1) The most common uropathogens are *Escherichia coli*. Other common causative organisms include *Enterococcus* spp., *Klebsiella* spp., *Proteus* spp., *Staphylococcus aureus* and *Staphylococcus saprophyticus* (2).

In Sri Lanka, nitrofurantoin, norfloxacin, cefuroxime, co-trimoxazole, and co-amoxiclav are recommended as empirical antibiotics for uncomplicated UTIs in adults. For complicated UTIs, such as UTIs in men/pyelonephritis, intravenous antibiotics like co-amoxiclav, ceftriaxone, ceftazidime, meropenem, or piperacillin-tazobactam are usually recommended (3).

A urine culture should be performed before commencing antibiotic treatment, and continuing antibiotic therapy should be based on an antibiotic sensitivity test (ABST) (4, 5). An ABST can identify the effective antibiotic(s) against specific uropathogens and guide the prescription of the most appropriate antibiotic (5,6).

Inappropriate antibiotic therapy has led to the emergence and spread of antimicrobial resistance (AMR) throughout the world, including in Sri Lanka (1) antibiotic susceptibility rates, association between (7).

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Though AMR is a concern in community-acquired UTIs, it is especially worrying in hospital settings where UTI is a common hospital-acquired infection(8). Although retrospective studies have described antibiotic sensitivity patterns of uropathogens in several outpatient settings in Sri Lanka. Less is known about the problem among hospitalized patients, particularly in Jaffna. (9,10).

Teaching Hospital Jaffna is the biggest tertiary care centre in the Northern Province serving a population of about 1.2 million. The hospital has a Microbiology Unit operating under a Consultant Microbiologist. Although healthcare facilities should have their own antibiotic policies based on the local sensitivity patterns (11). Such policies are yet to be developed for the Teaching Hospital Jaffna.

This study aims to describe the uropathogens isolated, their antibiotic sensitivity patterns and associated factors among adult inpatients (≥ 18 years) with a positive urine culture investigated at the Microbiology Unit of the Teaching Hospital Jaffna.

Methods

This institution-based cross-sectional study based on secondary data was carried out at the Microbiology Unit of Teaching Hospital Jaffna. Data were extracted from all positive urine culture reports and relevant request forms of adult inward patients (≥ 18 years) investigated at the Microbiology Unit between October 1, 2020 and December 31, 2020. Data were analyzed with the Statistical Package for Social Sciences (SPSS v27). Standard descriptive statistics were used to describe patient characteristics, uropathogens and antibiotic sensitivity patterns. The association between age and sex and the type of uropathogen was determined using chi square test with the critical level set at 0.05.

Ethics approval for this study was obtained from the Ethics Review Committee, Faculty of Medicine, University of Jaffna.

Results

Data were extracted from a total of 426 positive urine culture reports and relevant request forms. The mean age of the patients was 53.2 years (SD 19.9) with a median

of 58 years (IQR 35-70). Just under half the reports belonged to patients who were 60 years of age or above (47.2%, n=201), and the majority were females (60.1%, n=256; Table 1).

Table 1. Age and sex distribution of patients with a positive urine culture (n=426)

Demographic details		n	%
Age (years)	<40	132	31.0
	40-59	93	21.8
	≥ 60	201	47.2
Gender	Male	170	39.9
	Female	256	60.1
Total		426	100.0

Table 2 describes the uropathogens documented in positive urine culture reports of inpatients. Coliforms were the most common (63.4%, n=270) followed by *Candida* spp. (19.2%, n=82), *Pseudomonas* spp. (8.2%, n=35), *Acinetobacter* spp. (4.2%, n=18) and *Enterococcus* spp. (4.2%, n=18). *Staphylococcus aureus* was the least common (0.7%, n=3).

Table 2. Uropathogens isolated from positive urine cultures (n=426)

Uropathogen	n	%
<i>Coliforms</i>	270	63.4
<i>Candida</i> spp	82	19.2
<i>Pseudomonas</i> spp.	35	8.2
<i>Acinetobacter</i> spp.	18	4.2
<i>Enterococcus</i> spp.	18	4.2
<i>Staphylococcus aureus</i>	3	0.7
Total	426	100.0

Antibiotic therapy given before culture was documented in 183 (43%) inpatients. Details of prior antibiotic therapy were not documented in 36 request forms (8.5%) whereas the remainder (48.5%, n=207) had not received prior antibiotic therapy. Based on documented prior antibiotic therapy among 183 patients, the most commonly prescribed antibiotic was co-amoxiclav (24%, n=44), followed by ceftriaxone (19.7%, n=36), and ciprofloxacin (15.8%, n=29) (Figure 1).

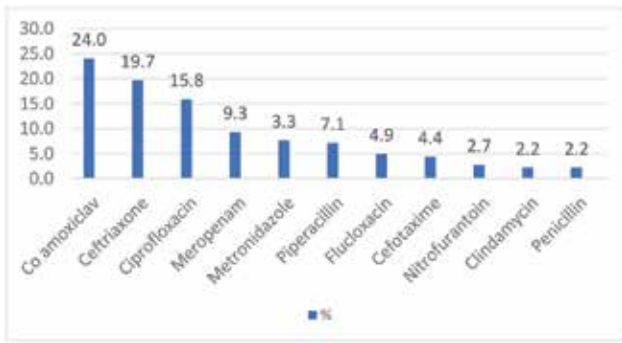


Figure 1. Antibiotic used prior to culture (n=183)

Our analysis of antibiotic sensitivity patterns showed that a large proportion of coliform isolates showed resistance to ampicillin (90.5%), ceftriaxone (60%), amoxicillin (41.1%), norfloxacin (42.8%), and ciprofloxacin (39.9%), while comparatively greater proportion of coliform isolates showed susceptibility to amikacin (83.1%), gentamicin (77.6%) and nitrofurantoin (76.3%). Meropenem resistance was seen in 9.3 % of the coliforms (Table 3).

Table 3. Antibiotic sensitivity patterns of uropathogens

Antibiotic	CF			PA			A			EC			C			SA		
	S	I	R	S	I	R	S	I	R	S	I	R	S	I	R	S	I	R
Meropenem	87.6	3.8	9.3	66.7	0	33.3	-	-	-	35	0	35	-	-	-	-	-	-
Amikacin	83.1	4.2	12.7	79	0	30.6	-	-	-	23.1	0	76.9	-	-	-	-	-	-
Gentamicin	77.6	1.6	20.8	69.7	0	30.3	-	-	-	44.6	0	55.4	-	-	-	-	-	-
Nitrofurantoin	76.3	10.5	13.2	-	-	-	88.9	0	11.1	0	0	100	100	0	0	-	-	-
Piperacillin	66.1	1.6	21.7	40	10	30.0	-	-	-	36.4	0	63.6	-	-	-	-	-	-
Tazobactam	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Ceftriaxone	58.1	0.6	40.9	56	0	30.0	0	0	100	37.1	14.3	32.6	100	0	0	-	-	-
Ciprofloxacin	58.3	1.6	39.9	67.6	0	32.4	30	0	30	33.3	0	66.7	0	30	30	-	-	-
Cefotaxime	39.3	1.7	39.2	-	-	-	-	-	-	0	0	100	-	-	-	-	-	-
Norfloxacin	36.4	3.8	42.5	67.7	0	32.3	18.2	18.2	63.6	28.6	0	71.4	0	0	100	-	-	-
Cefuroxime	33.9	2.3	43.3	-	-	-	-	-	-	33.3	0	66.7	-	-	-	-	-	-
Cephalexin	48.3	1.2	50.6	-	-	-	-	-	-	0	0	100	-	-	-	-	-	-
Amoxicillin	48.1	10.8	41.1	0	0	100	-	-	-	0	30	70	-	-	-	-	-	-
Nalidixic acid	44	0	56	100	0	0	30	0	30	66.7	0	33.3	-	-	-	-	-	-
Cotrimoxazole	40	0	60	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Ceftazidime	33.2	0	66.7	78.8	3	18.2	-	-	-	40	12.2	46.7	-	-	-	-	-	-
Acetaminophen	30.5	7.1	62.4	33.3	11.1	35.6	-	-	-	30.0	0	70	-	-	-	-	-	-
Taxolol	11.1	0	88.9	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Clavulanic acid	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Augmentin	9.3	0	90.7	-	-	-	33.3	0	66.7	-	-	-	0	0	100	-	-	-
Imipenem	0	0	100	66.7	0	33.3	-	-	-	36.4	0	63.6	-	-	-	100	0	0
Vancomycin	-	-	-	-	-	-	23.3	15.4	61.5	-	-	-	-	-	-	100	0	0

CF- Coliforms, PA- *Pseudomonas aeruginosa*, A-*Acinetobacter*, EC-*Enterococcus*, SA- *Staphylococcus aureus*, S- Sensitive, I- Intermediate, R- Resistant

A comparatively higher proportion of *Pseudomonas* isolates were sensitive to ceftazidime (78.8%), amikacin (70%) and gentamicin (69.7%), while only 40% were sensitive to piperacillin-tazobactam. Meropenem resistance was seen in 33.3% of the *Pseudomonas* isolates. Only 23.1% of *Acinetobacter* isolates were sensitive to amikacin and 36.4% to piperacillin-tazobactam. Meropenem sensitivity was seen in 75% of the *Acinetobacter* isolates. *Enterococcus* isolates showed good sensitivity to nitrofurantoin (88.9%) with a lower proportion showing sensitivity to ampicillin (38.9%) and norfloxacin (18.2%). Only 23.1% of the Enterococci were sensitive to vancomycin. *Staphylococcus aureus* showed 100% sensitivity to nitrofurantoin, cotrimoxazole and vancomycin, while 50 % of the isolates showed resistance to ciprofloxacin (Table 3).

Overall, 75.7% of all isolates were susceptible to nitrofurantoin. Overall susceptibility of uropathogens to ciprofloxacin was 57.8 % and to ceftriaxone 40% and meropenem 87.2%.

We found evidence of an association between the uropathogen isolated and age ($X^2=18.89$, $df=10$, p value=0.042) and sex ($X^2=12.35$, $df=5$, p value=0.030) of the patients (Table 4).

Table 4: Association of age and sex with uropathogen isolated

		n	CF	PA	A	EC	C	SA	p value
Age	<40	132	84 63.6%	12 9.1%	11 8.3%	4 3.0%	20 15.2%	1 0.8%	0.042
	40-59	93	63 67.7%	6 6.5%	4 4.3%	4 4.3%	14 15.1%	2 2.2%	
	≥60	201	123 61.2%	17 8.5%	3 1.5%	10 5.0%	48 23.9%	0 0%	
Sex	Male	170	102 60%	21 12.4%	3 1.8%	10 5.9%	33 19.4%	1 0.6%	0.03
	Fe-male	256	168 65.6%	14 5.5%	15 5.9%	8 3.1%	49 19.1%	2 0.8%	

CF- *Coliforms*, PA- *Pseudomonas aeruginosa*, A-*Acinetobacter*, EC-*Enterococcus*, C- *Candida* spp, SA- *Staphylococcus aureus*

Discussion

Among the organisms causing UTI in the present study, coliforms (63.5%) stand at the top of the list as has been previously described in several studies (1,2,6,13) followed by *Candida* spp., *Pseudomonas* spp., *Enterococcus* spp. (4.2%), *Acinetobacter* spp. and *Staphylococcus aureus*. The high percentage of *Candida* isolated in the present study indicates the need to consider whether they are an important cause of UTI among inpatients in our settings. This high proportion may be due to poorly controlled diabetes and/or immunocompromised status(12), given the high community prevalence of type 2 diabetes mellitus in Jaffna(13). However, another reason for *Candida* in urine could be contamination of urine samples due to indwelling catheters (14).

A substantial proportion of hospitalized patients with UTI had received antibiotics prior to culture, perhaps because the study focused on inpatients who are more likely to have been treated prior to admission. It is

noteworthy that whether antibiotics were given prior to culture was not documented in 8.5% of the request forms, signaling the need for improved investigation requisition practices in ward settings.

The most commonly prescribed antibiotic in the present study was co-amoxiclav, followed by ceftriaxone, ciprofloxacin and meropenem (Figure 1). It is encouraging to know that the most commonly prescribed antibiotic for inpatients with UTI was co-amoxiclav in our study as it is the recommended first-choice antibiotic for complicated UTIs and pyelonephritis. However, we could not describe its sensitivity pattern from the data available owing to the unavailability of co-amoxiclav discs at the Microbiology Unit during the period under study. Parenteral antibiotics such as ceftriaxone and meropenem may have been prescribed before culture to these patients because they had recurrent UTI with pathogens resistant to oral antibiotics or because they were treated for acute pyelonephritis/complicated UTI. This pattern of prescription differs from that described in a study among inpatients at Colombo North Teaching Hospital, where the most frequently prescribed empirical antimicrobial was ciprofloxacin (54.6%), followed by cefuroxime (12.3%) and nitrofurantoin (9.2%)(1). This difference might be due to the varying spectrum of clinical conditions for which antibiotics were prescribed in the two studies which we cannot confirm in our study as the indication for antibiotic prescription was not documented in the request forms.

The sensitivity pattern of coliforms to nitrofurantoin (76.3%) in our study was similar to that of a Colombo North Teaching Hospital study (74.8%), whereas a higher percentage of susceptibility was seen in India (90%) and in the United Kingdom (93.9%)(1). In the present study, coliforms had higher resistance to antibiotics such as co-trimoxazole (40.9%) and norfloxacin (42.8%), which can be used in the empirical treatment of uncomplicated cystitis than the Colombo North Teaching Hospital study (1). The large proportion of coliform isolates resistant to amoxicillin (41.1%), ceftriaxone (60%) and ceftazidime (66.7%)—the latter which is recommended in the treatment of complicated UTI and pyelonephritis(4)(5)—is comparable to that

of the Colombo North study(1). Though sensitivity of coliform isolates to ciprofloxacin, the third most commonly prescribed empirical antibiotic in the present study, was only 58%, susceptibility was notably greater than that reported in the Colombo North Teaching Hospital study where only 37.7% of coliform isolates were susceptible to ciprofloxacin(1). A higher proportion of coliforms were sensitive to amikacin in Colombo North Teaching Hospital study (93.1%) compared to that of ours (83.1%). However, gentamicin sensitivity to coliforms was higher (77.6%) in the present study than in the Colombo North Teaching Hospital study (63.0%).

We found a higher percentage of coliforms (9.3%) in the present study showed resistance to meropenem, an antibiotic that has been the ultimate option for several drug resistant uropathogens, including extended spectrum beta-lactamase (ESBL) producers. This result is similar to that of the study carried out at Colombo North Teaching Hospital (10.4%). According to the authors of the latter study, the ampicillin susceptibility reported in their study (13.4%) was one of the lowest reported in the literature (1). However, our study elicited an even lower susceptibility to ampicillin (9.5%).

More than 65% of the *Pseudomonas* isolates were sensitive to ciprofloxacin, gentamicin and ceftazidime in our study which is higher than the sensitivity pattern seen in the Colombo North study(1). although susceptibility to amikacin and meropenem was comparatively lower in our study. It is noteworthy that a high percentage of resistance (50%) was seen to piperacillin-tazobactam among *Pseudomonas* isolates and an even higher percentage of resistance (63.6%) to piperacillin-tazobactam among *Acinetobacter* isolates in the current study. Indeed, more than 60% of *Acinetobacter* in our study showed resistance to most of the antibiotics tested, including amikacin (76.9%) and a quarter were resistant to meropenem.

Enterococcus spp. showed higher resistance to norfloxacin (63.6%), vancomycin (61.5%) and ampicillin (61.1%) and good sensitivity to nitrofurantoin (88.9%). The high percentage of vancomycin-resistant *Enterococci* seen in our study is of great concern. It is much higher than the colonization seen in ICU patients

at the National Hospital of Sri Lanka in 2012 (18). All *Staphylococcus aureus* in our study were resistant to ciprofloxacin, whereas around 40% were sensitive in the Colombo North study.

According to a recent study in China by Wei Zhang *et al*, piperacillin-tazobactam is an effective, safe, and definite treatment option for complicated UTIs by ESBL-producing Enterobacteriaceae (19). However, piperacillin-tazobactam resistance was high among the uropathogens documented in our study, with resistance among *Pseudomonas spp*, 50%, *Acinetobacter spp.*, 63.6% and even coliforms, 31.7%. These findings question its use in the empirical treatment of complicated UTI in patients exposed to antibiotics or hospitalized recently.

In our study, *Pseudomonas* and *Enterococcus* isolates were seen more in male patients, while females had more *Acinetobacter*. *Candida* and *Enterococcus* were isolated more in the age group of more than 60 years, whereas *Acinetobacter* was isolated among the younger group of less than 40 years. These results are consistent with the Colombo North study, which found that *Pseudomonas* and *Candida* isolates were seen more in male patients and elderly patients, respectively (1). In a China study, *Pseudomonas*, *Enterococcus* and *Acinetobacter* isolates were seen more in female patients and *Enterococcus* and *Acinetobacter* displayed an age-related increase in prevalence.(20).

High resistance among the uropathogens seen in our study to most antibiotics tested, including empirical antibiotics, can lead to treatment failure and possible sepsis. Further, it will demand the use of more toxic and expensive antibiotics and more hospital admissions. These findings highlight the need for urgent measures to address antimicrobial resistance in the hospital setting. An initial step would be to develop and implement an institutional policy to ensure the collection of urine samples for culture prior to empirical therapy and to continue antibiotic therapy according to local sensitivity patterns(21). Of course, it is essential to ensure education and pre- and in-service training regarding the basics of urine collection and catheter care for healthcare professionals and adherence to infection prevention and control measures during urine collection

for culture (22). As irrational use of antibiotics is a major contributor to antibiotic resistance (23), measures to incentivise adherence to institutional guidelines on antibiotic therapy and strict infection control measures are needed.

This study has some limitations. We relied solely on the request forms and culture and ABST reports for data. Therefore, we were unable to find adequate clinical data and the type of UTI, which are likely to be important factors associated with the uropathogens isolated from inpatients with UTI. Further, although co-amoxiclav was the most commonly prescribed empirical antibiotic, we could not describe the sensitivity patterns of uropathogens to co-amoxiclav due to the unavailability of discs at the Microbiology Unit during the period under study.

Conclusion

A high proportion of isolates from adult inpatients at Teaching Hospital Jaffna were found to be resistant to several commonly prescribed antibiotics. These findings indicate the need for regular surveillance of uropathogens and their antibiotic sensitivity patterns, institutional policies to guide antibiotic prescription, and in-service training on strict infection prevention and control measures. Efforts should be made to perform co-amoxiclav sensitivity tests on uropathogens as it is the recommended first-choice antibiotic for complicated UTIs and pyelonephritis. The prevalence of vancomycin-resistant enterococci (VRE) in our setup should be studied further. Piperacillin-tazobactam should be used with caution as an empirical antibiotic for complicated UTI in our setting. A substantial proportion of hospitalized patients with UTI received antibiotics prior to culture, and a sizeable proportion of urine culture request forms did not contain details of prior antibiotic therapy. Therefore, improved investigation requisition practices in ward settings should be encouraged.

Acknowledgements

The authors would like to thank the staff in the Microbiology Lab who helped us in recruiting the culture request forms.

Conflicts of interest

The authors have no conflicts of interests to declare.

References

1. Wijekoon C, Dassanayake K, Pathmeswaran A. Antimicrobial susceptibility patterns and empirical prescribing practices in adult in patients with urinary tract infection: is there a need for changing clinical practices? *Sri Lankan J Infect Dis*. 2014;
2. Woldemariam HK, Geleta DA, Tulu KD, Aber NA, Legese MH, Fenta GM, et al. Common uropathogens and their antibiotic susceptibility pattern among diabetic patients. *BMC Infect Dis*. 2019;
3. Cheema S, Ur S, Cheema R. Prevalence of Antibiotic Resistance among Patients with Escherichia Coli Urinary Tract Infection in a Private Hospital at Lahore-Pakistan. 10(2).
4. Antibiotic Guidelines 2016 – Sri Lanka College of Microbiologists [Internet]. [cited 2024 May 23]. Available from: <https://slmicrobiology.lk/antibiotic-guidelines-2016/>
5. Kang CI, Kim J, Park DW, Kim BN, Ha US, Lee SJ, et al. Clinical Practice Guidelines for the Antibiotic Treatment of Community-Acquired Urinary Tract Infections. *Infect Chemother* [Internet]. 2018 Mar 1 [cited 2024 May 23];50(1):67–100. Available from: <https://pubmed.ncbi.nlm.nih.gov/29637759/>
6. Prakash D, Saxena RS. Distribution and antimicrobial susceptibility pattern of bacterial pathogens causing urinary tract infection in urban community of Meerut city, India. *ISRN Microbiol* [Internet]. 2013 Oct 29 [cited 2024 May 23];2013:1–13. Available from: <https://pubmed.ncbi.nlm.nih.gov/24288649/>
7. Fernando MMPS, Luke WANV, Miththinda JKND, Wickramasinghe RDSS, Sebastampillai BS, Gunathilake MPML, et al. Extended spectrum beta lactamase producing organisms causing urinary tract infections in Sri Lanka and their antibiotic susceptibility pattern -A hospital based cross sectional study. *BMC Infect Dis* [Internet]. 2017 Feb 10 [cited 2024 May 23];17(1). Available from: <https://pubmed.ncbi.nlm.nih.gov/28187754/>
8. Sedor J, Mulholland SG. Hospital-acquired urinary tract infections associated with the indwelling catheter. *Urol Clin North Am* [Internet]. 1999 [cited 2024 May 23];26(4):821–8. Available from: <https://pubmed.ncbi.nlm.nih.gov/10584622/>
9. Senadheera GP, Sri Ranganathan S, Patabendige G, Fernando GH, Gamage D, Maneke RM, et al. Resistance and utilisation pattern of antibacterial agents in outpatient settings in two Teaching Hospitals in Colombo. *Ceylon Med J* [Internet]. 2016 Sep 1 [cited 2024 Jun 28];61(3):113–7. Available from: <https://pubmed.ncbi.nlm.nih.gov/27727410/>
10. Jayatilleke SK, Patabendige G, Dassanayake M, Karunaratne GKD, Perera J, Perera RRD., et al. Analysis of urine culture isolates from seven laboratories of Sri Lanka: National Laboratory Based Surveillance of Sri Lanka College of Microbiologists in 2014. *Sri Lankan J Infect Dis* [Internet]. 2016 [cited 2024 Jun 28];6(1):17–24. Available from: <http://repository.kln.ac.lk/handle/123456789/12993>
11. Chew AB, Suda KJ, Patel UC, Fitzpatrick MA, Ramanathan S, Burns SP, et al. Long-term prescribing of nitrofurantoin for urinary tract infections (UTI) in veterans with spinal cord injury (SCI). *J Spinal Cord Med*. 2019 Jul;42(4):485–93.
12. Kojic EM, Darouiche RO. Candida infections of medical devices. *Clin Microbiol Rev* [Internet]. 2004 Apr [cited 2024 May 23];17(2):255–67. Available from: <https://pubmed.ncbi.nlm.nih.gov/15084500/>
13. Amarasinghe S, Balakumar S, Arasaratnam V. Prevalence and risk factors of diabetes mellitus among adults in Jaffna District. *Ceylon Med J* [Internet]. 2015 Sep 1 [cited 2024 May 23];60(3):107–10. Available from: <https://pubmed.ncbi.nlm.nih.gov/26520866/>
14. Singla N, Gulati N, Kaistha N, Chander J. Candida colonization in urine samples of ICU patients: determination of etiology, antifungal susceptibility testing and evaluation of associated risk factors. *Mycopathologia* [Internet]. 2012 Aug [cited 2024

- May 23];174(2):149–55. Available from: <https://pubmed.ncbi.nlm.nih.gov/22723047/>
15. Gautam G, Gogoi S, Saxena S, Kaur R, Dhakad MS. Nitrofurantoin Susceptibility Pattern in Gram-Negative Urinary Isolates: In Need of Increased Vigilance. *J Lab Physicians* [Internet]. 2021 Sep [cited 2024 May 23];13(3):252–6. Available from: <https://pubmed.ncbi.nlm.nih.gov/34602790/>
 16. Bean DC, Krahe D, Wareham DW. Antimicrobial resistance in community and nosocomial *Escherichia coli* urinary tract isolates, London 2005-2006. *Ann Clin Microbiol Antimicrob* [Internet]. 2008 Jun 18 [cited 2024 May 23];7. Available from: <https://pubmed.ncbi.nlm.nih.gov/18564430/>
 17. Wijekoon CN, Dassanayake K, Pathmeswaran A. Antimicrobial susceptibility patterns and empirical prescribing practices in adult in patients with urinary tract infection: is there a need for changing clinical practices? *Sri Lankan J Infect Dis*. 2014 Feb;4(1):9.
 18. Kannangara C, Chandrasiri P, Corea EM. Vancomycin resistant enterococcal (VRE) colonization among patients treated in intensive care units at the National Hospital of Sri Lanka, and determination of genotype/s responsible for resistance. *Ceylon Med J* [Internet]. 2018 Dec 31 [cited 2024 May 23];63(4):154. Available from: <https://pubmed.ncbi.nlm.nih.gov/30669209/>
 19. Zhang W, Yan CY, Li SR, Fan TT, Cao SS, Cui B, et al. Efficacy and safety of piperacillin-tazobactam compared with meropenem in treating complicated urinary tract infections including acute pyelonephritis due to extended-spectrum β -lactamase-producing Enterobacteriaceae. *Front Cell Infect Microbiol* [Internet]. 2023 [cited 2024 May 23];13. Available from: <https://pubmed.ncbi.nlm.nih.gov/37207190/>
 20. Zhan Z-S, Shi J, Zheng Z-S, Zhu X-X, Chen J, Zhou X-Y, et al. Epidemiological insights into seasonal, sex-specific and age-related distribution of bacterial pathogens in urinary tract infections. *Exp Ther Med* [Internet]. 2024 Feb 14 [cited 2024 May 23];27(4). Available from: <https://pubmed.ncbi.nlm.nih.gov/38476915/>
 21. Mori R, Lakhanpaul M, Verrier-Jones K. Diagnosis and management of urinary tract infection in children: summary of NICE guidance. *BMJ* [Internet]. 2007 Aug 25 [cited 2024 May 23];335(7616):395–7. Available from: <https://pubmed.ncbi.nlm.nih.gov/17717369/>
 22. Latour K, Lepeleire J De, Jans B, Buntinx F, Catry B. Diagnosis, prevention and control of urinary tract infections: a survey of routine practices in Belgian nursing homes. *J Infect Prev* [Internet]. 2020 Sep 1 [cited 2024 May 23];21(5):182–8. Available from: <https://pubmed.ncbi.nlm.nih.gov/33193820/>
 23. Chokshi A, Sifri Z, Cennimo D, Horng H. Global Contributors to Antibiotic Resistance. *J Glob Infect Dis*. 2019 Jan;11(1):36–42.